

MAXIMISING THE BENEFITS OF HERBICIDE RESISTANCE TESTING



Herbicide-resistant weeds are widespread in the UK but usually develop on individual fields as a consequence of the specific weed control programme used.

Resistance tests are a valuable management tool and this leaflet aims to identify when a test might be appropriate and to ensure sample collection, testing and interpretation are done correctly.

WHY HAVE WEED SEED OR PLANT SAMPLES FROM YOUR FIELDS TESTED?

- To know what you are dealing with – will herbicides work on **your** weeds?
- Avoid potentially wasteful use of herbicides
- Save money and prevent unnecessary use.
- Help assess the threat posed by resistance on newly farmed blocks of land.
- Monitor the success (or otherwise) of long-term resistance management strategies.

SEED SAMPLING FOR RESISTANCE TESTS

Most resistance tests rely on pot or Petri-dish assays using seed samples typically collected from weeds surviving herbicide treatment in the field. Some tests are also available using plants collected from the field.

HOWEVER...REMEMBER

- Resistance tests are only as good as the sample collected and the extent to which it represents the suspect population.
- Poor quality or immature seed = poor or no test result, which can be misleading and potentially worse than useless.
- Research has shown that there is negligible variation in resistance within an average black-grass patch, greater resistance variation between different patches within one field and lots of variation between different fields within one farm. One farm may therefore require different field samples.



HOW TO COLLECT A QUALITY SEED SAMPLE

- Collect samples when the majority of seeds are mature and 10-20% have already shed. Collecting too early or too late is likely to lead to samples with low viability.
- Do not simply collect intact heads, as many seeds on those will be immature.
- Ensure samples are representative of the problem field by collecting seeds from many plants in an area of about 100m by 2-3 tramlines, unless the problem occurs in a distinct, smaller patch
- For all species, allow seed to air-dry in a shallow open container for a few days and then transfer to paper envelopes for storage and transport (never store in polythene bags).
- Send samples to the testing centre as soon as possible after collection.
- Keep a record of both the weed infestation level and the area sampled (either a sketch map or a GPS reading). This information greatly enhances the value of any resistance test result.
- [Watch the video - Testing for resistance](#)

ITALIAN RYE-GRASS & WILD-OATS

In winter cereals collect seeds in late July/early August by holding heads or panicles inside a large polythene bag and shaking vigorously. Samples collected in the correct manner will consist almost entirely of seeds, with few stalks and little chaff or other debris. Aim to collect an amount equivalent to a mugful of seeds.



BLACK-GRASS

In winter cereals collect seeds in early July/early August. Never collect in June as that's too early. Collect seeds on dry days by gently rubbing heads over a bag or tray so only ripe seeds fall off.

Aim to collect an amount equivalent to a mugful of seeds if possible.



BROME

Sterile brome is the first grass to ripen in early June and the period for collection is over by the end of July. **Meadow** and **rye** brome are ready to collect from mid-late July through August. Collect seeds by gently stroking the heads over a bag or tray to remove the ripe seeds. Aim to collect a mugful.



BROAD-LEAVED WEEDS

Mature capsules of **poppies** can be collected in late July/August and seeds shaken out either in the field, or later. **Chickweed** plants can be pulled up when seeds start shedding in June/July and placed upside down in paper bags – seeds will fall out naturally. **Mayweed** heads should be collected when mature, in August, and seeds rubbed out later



THE BENEFITS AND LIMITATIONS OF RESISTANCE TESTS

- Seeds or plants collected for resistance tests usually represent a biased sample as they have (usually) been collected from survivors of herbicide treatment. Hence, resistance test results can exaggerate the level of resistance in the field, bearing in mind that there will be many older seeds in the soil seed bank which may produce plants that are still susceptible. Thus, **the detection of resistance does not mean that herbicides will necessarily fail in the field.**
- This bias in resistance testing should not be viewed as a limitation, but as a positive attribute, as confirmation of resistance provides an early warning sign of greater potential problems ahead. **Resistance rarely decreases** – it's a one-way ticket – so this early warning should be heeded and steps taken to reduce the resistance risk.
- Resistance test results **should be related to the sample location, the weed infestation and herbicide performance.** The relevance of the resistance test result in terms of its applicability to the entire field depends on the area sampled, the weed infestation and distribution, and the proportion of plants that survived treatment in the field. This information helps in assessing the likely impact of resistance on herbicide performance. The higher the weed density, the sooner major problems are likely if resistance has been confirmed.



RESISTANCE TESTING METHODS

There are several testing methods available and each has its advantages:

Glasshouse pot tests using seeds - Seeds are sown in pots and established plants are treated with herbicide (typically at 2-3 leaf stage for post-emergence applications) and assessed 3-4 weeks later;

Glasshouse pot tests using plants (e.g. Syngenta Quick Test) - Surviving plants are collected from the field, transferred to pots (after trimming), allowed to regrow, sprayed, assessed.

Petri-dish assays - Seeds are germinated in herbicide solutions and seedling growth recorded after 14 days.

Molecular tests - Leaves are tested in a laboratory for specific mutations known to confer target site resistance.

RISQ (Resistance In-season Quick Test) - Small to early tillered plants are collected from the field and placed in agar solution containing a herbicide dose and root growth is measured after 14 to 21 days.

Field-based diagnostic assay - Leaves from individual plants are extracted and analysed for the presence of a protein that regulates enhanced metabolism resistance (EMR).

Radio-labelled lab tests - Plants are collected from the field and the rate of metabolism of radio-labelled herbicide in leaves is measured.



SUMMARY OF RESISTANCE TEST METHODS

	Answer in same crop year	Suitable for all weed species	Mimics field conditions	Suitable for all herbicides	Detects resistance regardless of mechanism	Test duration	Cost
Glasshouse pots – seeds from field	No	Yes	Yes	Yes	Yes	Slow	Medium
Glasshouse pots – plants from field	Yes	Potentially	Yes	Most (not pre-em)	Yes	Medium	Medium
Petri-dish germination	No	No	No	No	No	Fast	Low
Molecular lab assays	Yes	Potentially	No	No (ACCase and ALS only)	No (TSR only)	Very fast	Medium-High
RISQ test	Yes	Potentially, but grasses best	No	Yes	Yes	Fast	Medium to high
Field-based diagnostic assay	Yes	Potentially	No	No	No (EMR only)	Very fast	Medium to High
Radio-labelled lab assays	Yes	Potentially	No	Potentially	No (EMR only)	Very Fast	Very High

ACCase: Acetyl CoA Carboxylase inhibitor herbicides (fops, dims & dens)

ALS: Acetolactate Synthase inhibitor herbicides

TSR: Target site resistance mechanism

EMR: Enhance metabolism resistance mechanism

INTERPRETATION OF TEST RESULTS

It is desirable that any resistance test:

- Includes susceptible and resistant reference populations. These are populations with known response to herbicides. Ideally their resistance mechanisms will be known. These helps validate the testing methodology and greatly help in the interpretation of results.
- Provides photos of pots or dishes for both the test and reference populations as these can make more impact than written results or 'R' ratings alone (See below).
- States the assessment method and basis on which resistance is assigned. This is particularly important where populations show marginal or partial resistance.
- Incorporates the 'R' resistance rating system which has been widely used in the UK to assist in interpreting the results of resistance tests and relating these to the likely impact on herbicide performance.

'R' RESISTANCE RATING EXPLAINED

RRR - Resistance confirmed, highly likely to reduce herbicide performance

RR - Resistance confirmed, probably reducing herbicide performance

R? - Early indications that resistance may be developing, possibly reducing herbicide performance

S - Susceptible

For a full explanation, see the WRAG website: <https://ahdb.org.uk/wrag>



This leaflet was updated by Lynn Tatnell (ADAS) based on the original document from 2012 on behalf of the UK Weed Resistance Action Group (WRAG)
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